Comprehensive haplotype-resolved view of genomic variation and methylation with long-read nanopore sequencing

Jean Monlong 14/03/2025





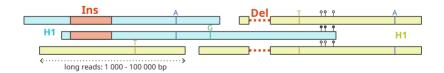
SYMPOSIUM FHU-G4 GÉNOMIQUE

Outline

Introduction: genomic variants, DNA methylation, long-read sequencing

Napu computational pipeline

Application to a cohort of rare disease patients



Introduction: genomic variants, DNA methylation, long-read sequencing

Different types of genomic variants

Single-nucleotide polymorphisms (SNPs)

polymorphisms
(INDELs)

Structural variants (SVs)

GATCAGC

GAT**CA**GC

GATCAGC

GAT**G**AGC

GAT - - GC

GATC_AGC

Different types of genomic variants

Single-nucleotide Insertion-deletion polymorphisms (SNPs)

polymorphisms (INDELs)

GATCAGC

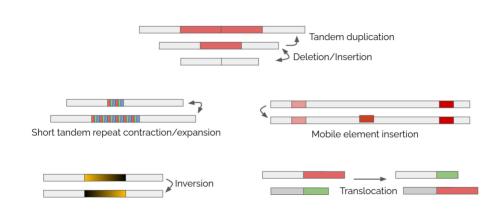
GATGAGC GAT - - GC

GATCAGC

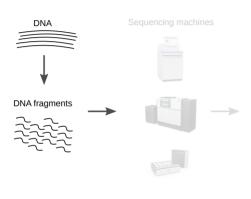
Different types of genomic variants

Single-nucleotide polymorphisms (SNPs)	Insertion-deletion polymorphisms (INDELs)	Structural variants (SVs)
GATCAGC	GAT CA GC	GATCAGC
GAT G AGC	GAT GC	GATC _▲ AGC
		CGC300bpGAT

Structural variants (SVs) come in diverse shapes and sizes



Genome sequencing



File (~100-300 Gb)

@ERR903030.219 HWI-D00574:82:C6L01ANXX:3:1101:3953:1913/1

AGCTCTTATTTTGAATATGTCCCATCAATACCTAATTTTTGGGAGGTTTTTAGCATGAAGGGTTGTTGAAT

GERR903030.220 HWI-0005748821C6L01AHXX:33:1101:3863:1914/1
ACCATGARACAGCAGTGTAGATCAGTACAAGAAGCACAGGGGGCATTGCATTTTGAGCATTTTGTATCA

9ERR903030.222 HWI-D00574:82:C6L01ANXX:3:1101:3833:1922/1

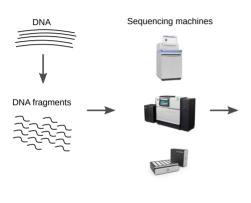
+

AAAAATGAACTAAAAATGCATTAAAGACCAAATGTAATACCTAAAAATGTAAAACTTTTAGAAGGAAACATAG

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Genome sequencing



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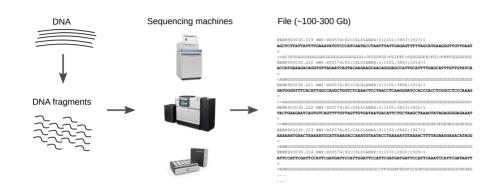
@ERR903030.219 HWI-D00574:82:C6L01ANXX:3:1101:3953:1913/1

 ${\tt GATGGGGTTCACATTGGCCAGGCTGGTCTCAACTCCTAACTCAAGGGATCCACCACCTCGGCCTCCCAAAG+}$

@ERR903030.222 HWI-D00574:82:C6L01ANXX:3:1101:3833:1922/1
TACTGAAGAATCAGTGTCAGTTTTGTTAGTTGTGATAATGACATTCTGCTAAGCTAAGTATAGAGGGGAGAAAT

@RRR903030.224 HMI-D00574:@2:C6L01ANXX:3:1101:3920:1929/1
ATTCCATTCGATTCGATTGGATTCCATTGGATTCCATTCGATGATGATTCCATTCAAATCCAT7
+

Genome sequencing



Sequencing reads

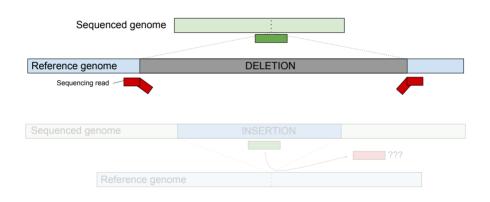
- 150-250 bp (current tech)
- ◆ 10,000s-100,000s bp (new tech. \$\$\$)

Aligning reads to a reference genome

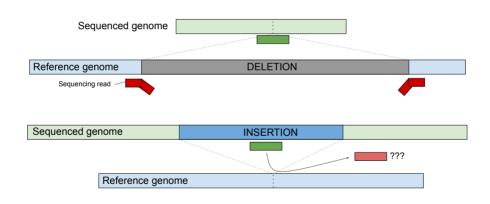


Assuming the reads are correctly placed, small variants are identified as recurrent differences between reads and the reference genome.

The challenges of structural variant detection



The challenges of structural variant detection



DNA methylation

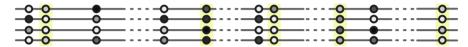
Epigenetic modification of the DNA, e.g. 5m-cytosine at CpG sites. More promoter methylation \rightarrow less transcription.



Aberrant methylation patterns can cause diseases (e.g. FMR1 in FXS).

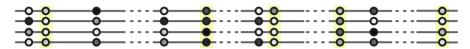
Episignatures of disease

Methylation pattern, across 10-100s of sites, associated with disease.

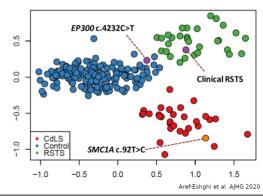


Episignatures of disease

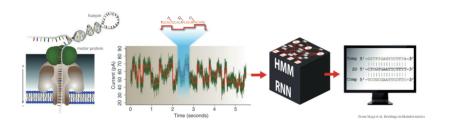
Methylation pattern, across 10-100s of sites, associated with disease.



Aref-Eshghi et al. (AJHG 2020) found an episignature with 34 genetic syndromes, from blood samples using methylation arrays.



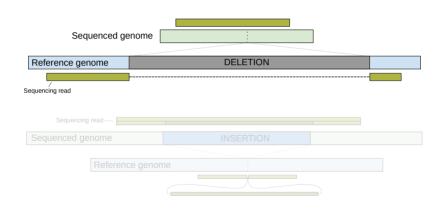
Long-read sequencing with Oxford Nanopore Technologies



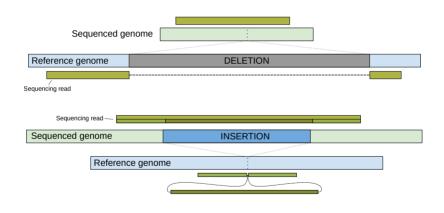
As the DNA (or RNA) fragment passes through the pore, the current changes and is decoded to predict nucleotides.

Reads length of 1,000s-100,000s of nucleotides.

Longer reads improve structural variant detection

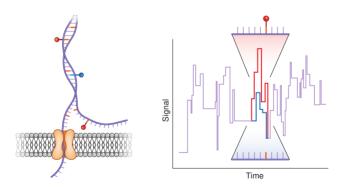


Longer reads improve structural variant detection



Nanopore sequencing can detect DNA/RNA modifications

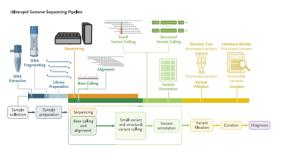
- 5-methylcysosine (5mC) for DNA/RNA
- 4-methylcysosine (4mC) for DNA
- N⁶-Methyladenine (6mA) for DNA/RNA



Schatz, Nature Methods 2023

ONT is portable (space!) and fast

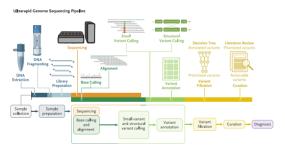
- Sequence as fast as possible
- Get a genomic diagnosis quick
- E.g. for newborns with suspicion of a rare genetic disease



Gorzynski et al. N. Engl. J. Med. 2022 Goenka, Gorzynski, Shafin, et al. Nat. Biotechnol. 2022

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"Fastest DNA sequencing technique": 5h2m



Napu computational pipeline

Cost-efficient Nanopore pipeline

- Only one flow-cell of Nanopore
- \sim ~30X coverage with 30 Kbp N50 reads

Cost-efficient Nanopore pipeline

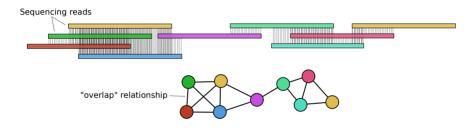
- Only one flow-cell of Nanopore
- ~30X coverage with 30 Kbp N50 reads
- Nanopore Analysis Pipeline (U?) to get haplotype resolved:
 - 1. small variants (SNPs/indels)
 - 2. structural variants
 - 3. de novo assembly
 - 4. methylation marks



Kolmogorov, Billingsley, et al. Nature Methods 2023

Longer reads enable *de novo* genome assembly

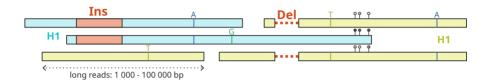
Reconstructs genomes without reference bias, hence better able to identify complex variants (e.g. combination of deletion/inversion)



The Shasta assembler is an overlap-layout-consensus assembler for Nanopore reads.

Shafin, Pesout, Lorig-Roach, Haukness, Olsen, et al. Nat. Biotechnol. 2020

Phased variants and methylation calls



Reads are **haplo-tagged** using information across heterozygous sites.

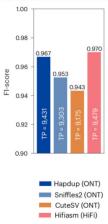
- Phased structural variants with Hapdup
- Phased small variants with DeepVariant
- Phased methylation calls with ModKit

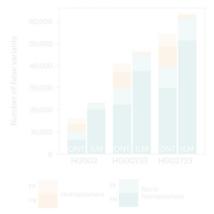
Kolmogorov, Billingsley, et al. Nature Methods 2023

Better calls for both small and structural variants...





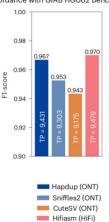




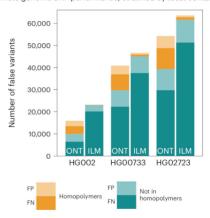
Kolmogorov, Billingslev, et al. Nature Methods 2023

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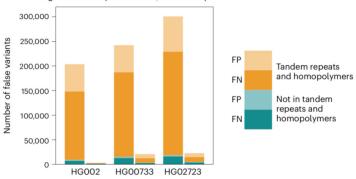
Whole genome SNP performance, stratified by local context



Kolmogorov, Billingsley, et al. Nature Methods 2023

...except for indels in homopolymers



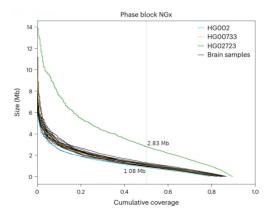


Note: Results above are for the R9 chemistry. The new R10 chemistry has lower error rate and better (indel) calling performance.

Kolmogorov, Billingsley, et al. Nature Methods 2023

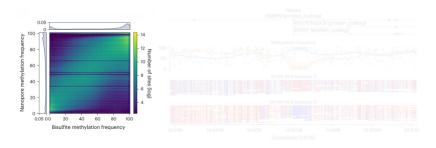
Variants and methylation phased in Mbp-long blocks

Small variants, structural variants, methylation marks are homogenized into megabase-long phase blocks.



Haplotype-resolved methylation at CpG sites

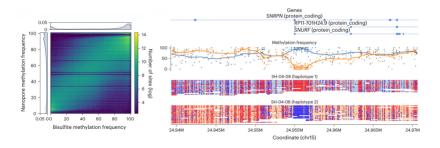
- Good concordance with short-read bisulfite sequencing.
- Haplotype-specific methylation patterns.



Kolmogorov, Billingsley, et al. Nature Methods 2023

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Kolmogorov, Billingsley, et al. Nature Methods 2023



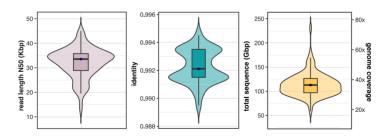
Application to a cohort of rare disease patients

Chan Zuckerberg Initiative®





42 probands and 56 unaffected family members, sequenced with one-flowcell of ONT long-read sequencing (R10).

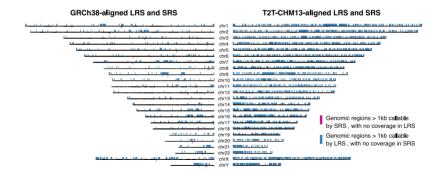


Negi et al. AJHG 2025

Better coverage of confidently mapped reads

More of the CHM13-T2T genome covered with at least 10x.

• 93.99% (LRS) vs. 88.27% (SRS)



Negi et al. AJHG 2025

Small variants found by long-reads only

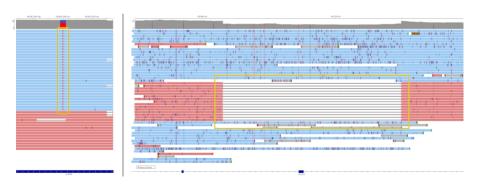
Missense mutation in *KRT86* disease gene (monilethrix) invisible with short reads.



Compound heterozygous variants thanks to phasing information

In *LHCGR* gene, associated with Leydig cell hypoplasia:

- Coding SNV on haplotype 1 (left, blue reads)
- ~7 Kbp deletion of an exon on haplotype 2 (right, red reads)

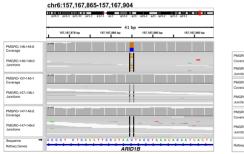


Negi et al. AJHG 2025

Patient with complex neurodevelopmental phenotype

Variant of Uncertain Significance SNV in *ARID1B* gene (Coffin-Siris syndrome 1?).

• *De novo*, SRS and LRS, new splice site predicted *in silico* (SpliceAI).

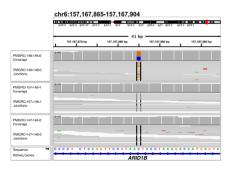


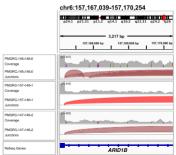


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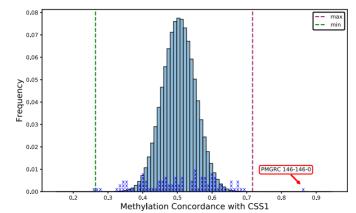
Negi et al. AJHG 2025

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Large-scale study of brain samples

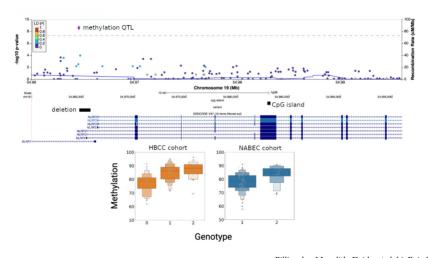
New resource: 351 brain control samples sequenced in the NIH's Center of Alzheimer's and Related Dementias (CARD) long-read sequencing initiative.

- 234,905 SVs
- >800 SV expression QTLs
- >2000 SV methylation QTLs



Billingsley, Meredith, Daida, et al. bioRxiv 2024

Example of a SV methylation QTL



Billingsley, Meredith, Daida, et al. bioRxiv 2024

Take-home message

Cost-effective **long-read sequencing** using nanopore technologies to help solve undiagnosed **rare disease** cases.



Haplotype-resolved

- small variants (SNPs/indels)
- structural variants
- de novo assembly
- methylation marks

Kolmogorov, Billingsley, et al. Nature Methods 2023

Negi et al. AJHG 2025

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Research Institute

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- Paolo Canigiula



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